

COLLECTION CATALOGUE

Catalogue of type specimens of the families Ceraphronidae, Platygastriidae, Proctotrupidae and Scelionidae (Hymenoptera) deposited in the National Museum of the Czech Republic

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Accepted:
15th December 2025

Published online:
31st December 2025

Abstract. Type specimens from the Hymenoptera collection deposited in the Department of Entomology, National Museum of the Czech Republic are currently being catalogued. In this part of the catalogue, we deal with the families Ceraphronidae, Platygastriidae, Proctotrupidae and Scelionidae. We present precise information about the 28 species belonging to the families Ceraphronidae (three species), Platygastriidae (three species), Proctotrupidae (two species) and Scelionidae (20 species), including six holotype specimens. For each species, we give the current status, distribution, and photographs, with many of these images being published here for the first time. In addition, two paratype specimens previously assigned to *Ceratoteleia leucosia* Nixon, 1931 and *Paratrimorus pinguis* Nixon, 1933 are herein revised and the species are transferred to the genera *Dicroscelio* Kieffer, 1913 and *Psilanteris* Kieffer, 1916, respectively, resulting in the following new combinations – *Dicroscelio leucosia* (Nixon, 1931) comb. nov. and *Psilanteris pinguis* (Nixon, 1933) comb. nov.

Key words. Hymenoptera, Ceraphronidae, Platygastriidae, Proctotrupidae, Scelionidae, catalogue, new combination, type specimens, National Museum of the Czech Republic

Zoobank: <http://zoobank.org/urn:lsid:zoobank.org:pub:0A2885F3-C351-43C9-B5C4-376791B0D50B>

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Introduction

The entomological collection of the National Museum of the Czech Republic (NMPC) houses over 8 million specimens, including tens of thousands of primary types. While most of these specimens, and the majority of types, belong to the order Coleoptera (beetles), the collection encompasses nearly all insect orders to varying extents. The Hymenoptera collection, estimated at around 550,000 specimens, represents a smaller but significant part of the collection with many primary types that are now catalogued in ongoing contributions. The history of the entomological collections in NMPC was previously reviewed for Coleoptera (BEZDĚK & HÁJEK 2009), Hemiptera (KMENT

& KOLÍNOVÁ 2013, MALENOVSKÝ et al. 2016), Diptera (TKOČ et al. 2014) and Polyneoptera (MACHÁČKOVÁ & FIKÁČEK 2014).

To date, the Hymenoptera families Formicidae, Bracnidae and superfamily Diaprioidea have been catalogued in the NMPC (BEZDĚČKOVÁ et al. 2017; HOVORKA & MACEK 2022, 2024). In this contribution, we continue this effort by providing a list of types, accompanied by images, for the families Ceraphronidae, Platygastriidae, Proctotrupidae and Scelionidae.

Ceraphronidae is a diverse, cosmopolitan family of parasitoid wasps within Ceraphronoidea, currently comprising over 600 described species. They are minute wasps with reduced wing venation and uniform external morphology, which makes their taxonomy difficult. Most species are



*) Catalogue of type specimens in NMPC, Part 20

parasitoids or hyperparasitoids of various insect hosts, although their biology is still poorly known (MIKÓ et al. 2016, TRIETSCH et al. 2018).

Platygastriidae is a moderately large family of parasitic wasps comprising 1,153 described species worldwide (AUSTIN et al. 2005). Together with Scelionidae, it belongs to the superfamily Platygastroidea (AUSTIN et al. 2005, MURPHY et al. 2007). Most species are endoparasitoids of Diptera (Cecidomyiidae), Hemiptera (Flatidae eggs, Pseudococcidae nymphs, Aleyrodidae nymphs), Hymenoptera (Crabronidae larvae), as well as some Coleoptera (Curculionidae and Cerambycidae) (MASNER 1993a, AUSTIN et al. 2005, CHEN et al. 2021). Recent phylogenetic studies confirmed the monophyly of the family and separated it from Scelionidae (CHEN et al. 2021).

Scelionidae is a diverse family of parasitic wasps comprising 3,308 described species worldwide (AUSTIN et al. 2005). They are specialized in parasitizing the eggs of insects and spiders (MASNER 1993b, AUSTIN et al. 2005). These minute wasps are often used as biological control agents, especially against hemipteran and lepidopteran pests (ORR 1998, AUSTIN et al. 2005). Phylogenetic analyses recently re-established Scelionidae as a distinct family within Platygastroidea (CHEN et al. 2021).

Proctotrupidae is the principal extant family of Proctotrupoidea, including about 650 described species in ca. 30 genera worldwide (IZADIZADEH et al. 2022, CAN 2023). Adults are relatively larger parasitoids, often with long ovipositors or wingless females adapted to a subterranean lifestyle. They are mainly koinobiont endoparasitoids of coleopteran larvae but some species attack also centipedes, dipteran and lepidopteran larvae (TOWNES & TOWNES 1981, IZADIZADEH et al. 2022, CAN 2023).

By documenting and facilitating access to type material, the catalogue aims to support taxonomic research and provide a stable foundation for nomenclature within these groups. This effort is anticipated to advance the understanding of parasitoid wasp diversity and the ecological roles these insects play across various ecosystems, furthering both taxonomic clarity and biodiversity conservation (TRIAPITSYN 2014, ZHANG et al. 2025).

Material and methods

The system used to arrange the taxa mostly follows JOHNSON (1992). Within each subfamily, the genera and species are arranged alphabetically. Each entry includes: – the name of the taxon in its original combination; – the name of the taxon in its original combination and spelling, with the author and year of description, along with pagination, figures, and plates; – the name-bearing type, number of specimens (including their sex if known) and exact label data. Our remarks are found in square brackets: [p] – preceding data are printed, [hw] – preceding data are handwritten. Separate labels are indicated by a double slash ‘//’ and lines within each label are separated by a slash ‘/’. Unless otherwise indicated, it means black ink and white label; – the type condition for considerably damaged specimens

(given in parentheses following the particular specimen); – the current taxonomic status; – known distribution data (when country-level data are unavailable, distribution is given at the biogeographical region level).

Photographs of type specimens and their labels were taken using an Olympus SZX16 stereomicroscope equipped with an Olympus DP73 camera, as well as a Leica M205C stereomicroscope fitted with a Canon R7 camera. Individual photographs were stacked using Helicon Focus 8.2.18 software and subsequently edited using Adobe Photoshop. The full reference to each cited publication can be found in the References section.

Catalogue

Family Ceraphronidae Haliday, 1833

Aphanogmus annulicornis Jansson, 1957

(Fig. 1)

Aphanogmus annulicornis Jansson, 1957: 71–74, fig. 1 (original description).

One paratype is deposited in NMPC (type collection):

PARATYPE (♀): ‘Ör. Ad. Lay [hw, indistinct characters] / 27/85 4 [hw] l.i. [hw, indistinct characters] // Paratypus [p, red label in black frame] // 9. [hw] // *Aphanogmus* / *annulicornis* ♀ / nov. sp. Jann. / in litt. [hw, pencil] / 26706 [hw] / ex coll. J. Straka / National Museum / Prague, Czech Republic [p, opposite side of the same label]’ (Fig. 1).

Current status. Junior synonym of *Aphanogmus compressus* (Ratzeburg, 1852) (DESSART 1991a).

Distribution. Palaearctic, known from Denmark, Finland, Germany, Ireland, Norway, Sweden, United Kingdom (DESSART 1991a).

Elysoceraphron hungaricus Szélenyi, 1936

(Fig. 2)

Elysoceraphron hungaricus Szélenyi, 1936: 64–66, figs 16, 17 (original description).

The allotype is deposited in NMPC (general collection):

ALLOTYPE (♂): ‘Bohemia sept. / Bělá u Děčína / 20. 8. 56 Bčk. [hw]’ (Fig. 2).

Current status. Valid species (BELOKOBYSKIJ et al. 2019).

Distribution. Palaearctic (BELOKOBYSKIJ et al. 2019), described from Hungary (SZÉLENYI 1936).

Sinaris roubali Gregor, 1939

(Fig. 3)

Sinaris roubali Gregor, 1939: 132–133, fig. 1 (original description).

The holotype is deposited in NMPC (general collection):

HOLOTYPE (♀): ‘Boh.-Hr. Králové / 18.iv.1938 / leg. J. Roubal [hw] / “Čeper- / ka [hw, vertically written] / “čeperka” [hw, reverse of the same label] // TYPUS [p, red label in black frame] // *Synarsis* / *roubali* / ♀ spn. [hw] / Det. Fr. Gregor [p]’ (Fig. 3). [The head is separated from the body but mounted on the top of the same label.]

Current status. Junior synonym of *Synarsis xanthothorax* Szélenyi, 1936 (DESSART 1991b).

Distribution. Palaearctic (DESSART 1991b), described from the Czech Republic (GREGOR 1939).

Family Platygastriidae Haliday, 1833

Subfamily Platygastriinae Haliday, 1833***Inostemma staryi* Masner, 1955**

(Fig. 4)

Inostemma staryi Masner, 1955: 140–142, fig. 1 (original description).

The holotype is deposited in NMPC (type collection):

HOLOTYPE (♀): ‘Bohemia c. / Karlštejn / P. Starý lgt. [hw] / 24.iv. / 1951 [hw, reverse side of the same label] // Mus. Nat. Pragae / Inv. [p] 3102 [hw, orange label] // *Inostemma / staryi* n.sp. [hw, red label]’ (Fig. 4).**Current status.** Valid species (MASNER & HUGGERT 1989).**Distribution.** Palaearctic, known from the Czech Republic (MASNER 1955).**Subfamily Sceliotrachelinae Brues, 1908*****Fidiobia hofferi* Kozlov, 1978**

(Fig. 5)

Fidiobia hofferi Kozlov, 1978: 656 (original description).

The lectotype (designated by POPOVICI & BUHL 2010) is deposited in NMPC (general collection):

LECTOTYPE (♀): ‘Moravia m. / Bzenec / Lemarie lgt. [hw] // 1.vii.58. / ex larva / Ichneumonid [hw] // Lectotypus [p, red label] // *Fidiobia / hofferi* Kozlov / det. O. Popovici, 2009 [p]’ (Fig. 5).**Current status.** Valid species, paralectotypes also deposited in Muséum National d’Histoire Naturelle, Paris (KOZLOV 1978, NOTTON 2010, POPOVICI & BUHL 2010).**Distribution.** Palaearctic, known from the Czech Republic, Finland, Iran, Romania, Sweden and Ukraine (KOPONEN & HUGGERT 1982, POPOVICI & BUHL 2010, ASADI-FARFAR et al. 2020, POPOVICI et al. 2022).***Iphitrachelus gracilis* Masner, 1957**

(Fig. 6)

Iphitrachelus gracilis Masner, 1957: 58–60, figs 2, 4, 8 (original description).

The holotype is deposited in NMPC (type collection):

HOLOTYPE (♂): ‘Slovakia c. / Ban. Štiavnica / Masner lgt [hw] / Lesní potok / 10.viii.55 [hw, reverse side of the same label] // *Iphitrachelus / gracilis* Masner / Holotypus [hw, blue ink] / L. Masner det., 19 [p] 57 [hw, blue ink] // Mus. Nat. Pragae / Inv. [p] 3104 [hw]’ (Fig. 6).**Current status.** Valid in its original combination (VLUG 1995).**Distribution.** Palaearctic, known from Austria, Czech Republic and Slovakia (OHIO STATE UNIVERSITY 2025).**Family Proctotrupidae Latreille, 1802*****Serphus gravidator antennalis* Tomšík, 1944**

(Fig. 7)

Serphus gravidator antennalis Tomšík, 1944: 51 (original description).

The holotype is deposited in NMPC (type collection):

HOLOTYPE (♂): ‘Somotor 2.viii [hw] / 193[9]6[hw] A. Hoffer [p] // ♂ [hw, red ink] // TYPUS [p, red label] // *Serphus / gravidator / antennalis / Tomšík, 1944* [hw, blue ink] // Mus. Nat. Pragae / Inv. [p] 25.511 [hw, blue ink, orange label]’ (Fig. 7).**Current status.** Junior synonym of *Proctotrupes gravidator* (Linnaeus, 1758) (JOHNSON 1992).**Distribution.** Holarctic, known from Austria, Belgium,

Canada, China, Czech Republic, Croatia, Denmark, England, Finland, France, Germany, Hungary, Ireland, Italy, Japan, Jordan, Mongolia, Norway, Poland, Romania, Russia, Slovakia, Slovenia, Spain, Sweden, Switzerland, Turkey, USA, and Iran (TOWNES & TOWNES 1981, IZADIZADEH et al. 2016).

***Serphus gravidator niger* Tomšík, 1944**

(Fig. 8)

Serphus gravidator niger Tomšík, 1944: 51 (original description).

Twelve syntypes are deposited in NMPC (type collection):

SYNTYPE (♀): ‘ČSR Moravia / Pouzdřany [p] 11.viii [hw] / Fr. Gregor 193[p]6[hw] // TYPUS [p, red label in black frame] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / LECTOTYPUS [p] ♀ [hw] / Bouček det. 196[p]3[hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.512 [hw, blue ink, orange label]’ (Fig. 8).SYNTYPE (♂): ‘ČSR Moravia / Pouzdřany [p] 14.ix. [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.513 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia / Fr Gregor 19 [p] / Bzenec 7.viii [hw, reverse side of the same label] // “empty red label” // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.514 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia / Ubuín [p] 31.v. [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.515 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia / Brno [p] Pal. v. 23.vii [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.516 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia / Brno [p] Jundr. 26.v. [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.517 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia [p] / Babice 7.viii. / 38 [hw] / Fr. Gregor 19 [p] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.518 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia [p] / Tišnov-Sekoř / 20.v. [hw] / 193[p]6[hw] A. Hoffer [p] // *Serphus gravidator / niger* Tomšík, 1944 [hw, blue ink] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw, red label] // Mus. Nat. Pragae / Inv. [p] 25.519 [hw, blue ink, orange label]’.SYNTYPE (♂): ‘ČSR Moravia / Fr. Gregor 19 [p] / Bzenec 7.viii.36 [hw, reverse side of the same label] // *Serphus gravidator / niger* Tomšík, 1944 / Paralectotypus (Běk)’.SYNTYPE (♂): ‘ČSR Moravia [p] / 24 / Čejč vii 38 [hw] / Fr. Gregor 19 [p] // *Serphus gravidator / niger* Tomšík, 1944 [hw] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw]’.SYNTYPE (♂): ‘ČSR Moravia / Brno [p] P.v.23.vii [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw]’.SYNTYPE (♂): ‘ČSR Moravia / Brno [p] Pal.23.vii [hw] / Fr. Gregor 193[p]6[hw] // *Serphus gravidator / niger* Tomšík, 1944 [hw] / Paralectotypus / Bouček det. 196[p]3 ♂ [hw]’.**Current status.** Junior synonym of *Proctotrupes gravidator* (Linnaeus, 1758) (JOHNSON 1992).**Note.** TOMŠÍK (1944) described *Serphus gravidator niger* based on twelve syntypes, eleven from Moravia: ‘Auerschitz [= Pouzdřany] 1 ♀, 1 ♂, Tschetsch [= Čejč] 1 ♂, Bisenz [= Bzenec] 1 ♂, Babitz [= Babice] 1 ♂, Brünn [= Brno] 4 ♂, Gross-Ubuschin [= Ubušín] 1 ♂ (Greg. [= Gregor]), Tischnowitz [= Tišnov] 1 ♂ (Hof. [= Hoffer])’ and one from Bohemia: ‘Sobieslau [= Soběslav] 1 ♂

(Tmš. [= Tomšík]). The latter syntype from Soběslav is missing from NMPC, the remaining eleven specimens from Moravia are present, but there are two identical male syntypes from Bzenec, both collected by F. Gregor on 7.viii.1936. It is impossible to distinguish which of the two specimens from Bzenec TOMŠÍK (1944) studied, or both, and the number of specimens given in the paper is a simple mistake. Therefore, both specimens are accepted as syntypes. Bouček intended to designate a lectotype and paralectotypes, indicated by labels he attached to the syntype series (Fig. 8). However, these designations were never published and the specimens retain their status as syntypes. **Distribution.** Holarctic, known from Austria, Belgium, Canada, China, Czech Republic, Croatia, Denmark, England, Finland, France, Germany, Hungary, Ireland, Italy, Japan, Jordan, Mongolia, Norway, Poland, Romania, Russia, Slovakia, Slovenia, Spain, Sweden, Switzerland, Turkey, USA, and Iran (TOWNES & TOWNES 1981, IZADIZADEH et al. 2016).

Family Scelionidae Haliday, 1839
Subfamily Scelioninae Haliday, 1939

***Breviscelio crenatus* Sundholm, 1970**

(Fig. 9)

Breviscelio crenatus Sundholm, 1970: 383 (original description).

One paratype is deposited in NMPC (general collection):

PARATYPE (♀): 'S. Afr. Cape Prov / Hermanus / 20.XII.50 No. 92 [p] // Paratypus [p] / Encyrtoscelio / crenatus / Sundh. [hw, red label]' (Fig. 9).

Current status. Valid as *Gryon crenatum* (Sundholm, 1970) (TALAMAS et al. 2021).

Distribution. Afrotropical, known from Botswana, Malawi, Namibia, South Africa and Zimbabwe (CALECA 1992).

***Ceratoteleia leucosia* Nixon, 1931**

(Fig. 10)

Ceratoteleia leucosia Nixon, 1931: 363–365, fig. 3 (original description).

Dicroscelio leucosius (Nixon, 1931), **comb. nov.**

One paratype is deposited in NMPC (type collection):

PARATYPE (♂): 'Para- / type [p, rounded label with yellow margins] // Port St. John, / Pondoland. / 1-17.Mar.1924. [p] // S. Africa. / R. E. Turner. / Brit. Mus. / 1924–177. [p] // *Ceratoteleia / leucosia*, Nixon. / Paratype 1931. [p] // *Dicroscelio leucosius* (Nixon, 1931) / Talamas E. det. 2025 [p]' (Fig. 10).

Generic transfer. MASNER (1965) transferred *Ceratoteleia leucosia* to *Calotelea* Westwood, 1837. This species was listed under *Calliscelio* Ashmead, 1893 in JOHNSON (1992) but without an explicit mention of a generic transfer. Most likely this was a simple error of name placement given the alphabetical proximity of *Calotelea* and *Calliscelio*. Examination of the paratype specimen in NMPC reveals that it belongs in neither of these genera. The robust malar striae (Fig 10) exclude it from *Calliscelio*, in which these striae are entirely absent (TALAMAS et al. 2016). *Calotelea* is characterized by the presence of a skaphion, a long radicle, a thin lamella on the metascutellum (variable in shape), a slender, gracile form without coarse sculpture, and seven externally visible segments of the metasoma.

Rarely, species may have the skaphion absent, usually in males, and there is some variability in the length of the radicle. However, *Ceratoteleia leucosia* is incongruent with each of these characters. We place this species in *Dicroscelio* Kieffer, 1913, which is commonly found throughout Africa, based on the broad, rugulose armature of the metascutellum, the long T3/S3 on the metasoma, and venation of the forewing: the marginal vein is short and the postmarginal and stigmal veins are approximately equal in length. The limits of *Dicroscelio* are not well understood. It is cosmopolitan and somewhat polytypic, but *Dicroscelio leucosia* and the type species, *D. flavipes* (Kieffer, 1913), have very similar shapes of the metascutellum and a dip in the submarginal vein, suggesting a close relationship.

Current status. Valid as *Dicroscelio leucosius* (Nixon, 1931) **comb. nov.**

Distribution. Afrotropical, known from the type locality in South Africa (NIXON 1931).

***Duta tenuicornis fragilis* Nixon, 1933**

(Fig. 11)

Duta tenuicornis fragilis Nixon, 1933: 306–309, text fig. 4 (original description).

One paratype is deposited in NMPC (type collection):

PARATYPE (♂): 'Para- / type [p, rounded label with yellow margins] // Port St. John, / Pondoland. / Jan. 1924. [p] // S. Africa. / R.E. Turner. / Brit. Mus. / 1924–97. [p] // *Duta / tenuicornis / Dodd. / ssp. fragilis* Nixon / Paratype ♂, 1933 [hw]' (Fig. 11).

Current status. Junior synonym of *Duta tenuicornis* (Dodd, 1920) (NIXON 1933).

Distribution. In addition to the type locality in South Africa, this species has been reported from Russia, Azerbaijan, Japan, Greece, South Africa, Indonesia, Moldova, Ukraine, Romania (POPOVICI 2005, OHIO STATE UNIVERSITY 2025).

***Echthrodesis lamoralis* Masner, 1968**

(Fig. 12)

Echthrodesis lamoralis Masner, 1968: 198, figs 1–5 (original description).

Two paratypes are deposited in NMPC (general collection):

PARATYPE (♀): 'From eggs of / *D. fomidabilis* / April 1966 [hw] // South Africa Cape / Kommetiie "The Island" / B. Lamoral colle. [hw] // Paratype [hw, pink label] // *Echthrodesis / lamoralis* Msn. / ♀ [hw] / L. Masner det., 19[p]68[hw]' (Fig. 12).

PARATYPE (♀): 'From eggs of / *D. fomidabilis* / April 1966 [hw] // South Africa Cape / Kommetiie "The Island" / B. Lamoral colle. [hw] // Paratype [hw, pink label] // *Echthrodesis / lamoralis* Msn. / ♀ [hw] / L. Masner det., 19[p]68[hw]'.

Current status. Valid species (MASNER 1968, VAN NOORT et al. 2014, OWEN et al. 2014)

Distribution. Afrotropical, known from Cape Peninsula, South Africa (MASNER 1968, VAN NOORT et al. 2014, OWEN et al. 2014).

***Govinda nona* Nixon, 1933**

(Fig. 13)

Govinda nona Nixon, 1933: 466–467, text fig. 8 (original description).

One paratype is deposited in NMPC (type collection):

PARATYPE (♀): 'Para- / type [p, rounded label with yellow margins] // Mossel Bay, / Cape Province. / April, 1921. [p] // S. Africa. /

R.E.Turner. / Brit. Mus. / 1921–210. [p] // *Govinda / nona* Nixon / Paratype 1933. [p] (Fig. 13).

Current status. Valid as *Dicroscelio nonus* (Nixon, 1933) (YODER et al. 2009).

Distribution. Afrotropical, known from South Africa (VAN NOORT 2025, OHIO STATE UNIVERSITY 2025).

Hadronotus lymantriae Masner, 1958

(Fig. 14)

Hadronotus lymantriae Masner, 1958: 39–43, fig 1 (original description).

The holotype and two paratypes are deposited in NMPC (general collection):

HOLOTYPE (♀): ‘Slovakia mer. / Beluja 25.iii.1965 / Ing. M. Čapek lgt. [hw, blue ink] // *Lymantria / dispar* L. / ova [hw, blue ink] // *Hadronotus* ♀ / *lymantriae* n.sp. / (Entomophaga 3,1958) [hw, blue ink] // Holotypus [hw, red label]’ (Fig. 14).

PARATYPES (2 ♀♀): ‘Slovakia mer. / Beluja 25.iii.1955 / Ing. M. Čapek lgt. [hw, blue ink] // *Lymantria / dispar* L. / ova [hw, blue ink] // *Hadronotus* ♀ / *lymantriae* n.sp. / (Entomophaga 3,1958) [hw, blue ink] // Paratypus [hw, red label]’.

Current status. Valid as *Gryon lymantriae* (Masner, 1958) (MASNER 1965, TALAMAS et al. 2021).

Distribution. Palearctic, known from Azerbaijan, Bulgaria, France, Greece, Iran, Italy, Moldova, Slovakia, Tajikistan, Ukraine and Uzbekistan (MASNER 1958, JOHNSON 1992, OHIO STATE UNIVERSITY 2025).

Macroteleia unicolor Nixon, 1931

(Fig. 15)

Macroteleia unicolor Nixon, 1931: 371–374, fig. 5 (original description).

One paratype is deposited in NMPC (general collection):

PARATYPE (♀): ‘Para- / type [p, round label in yellow frame] // Port St. John, / Pondoland. / 6-25.Feb.1924. [p] // S. Africa. / R.E.Turner / Brit. Mus. / 1924–136. [p] // *Macroteleia / unicolor*, Nixon. / Paratype 1931. [p]’ (Fig. 15).

Current status. Preoccupied, valid under the replacement name *Macroteleia nixon* Masner, 1965 (MASNER 1965)

Distribution. Afrotropical, known from the type locality in South Africa (NIXON 1931).

Nardo cumaeus Nixon, 1938

(Fig. 16)

Nardo cumaeus Nixon, 1938: 279 (original description).

Three paratypes are deposited in the NMPC (type collection):

PARATYPES (3 ♀♀): ‘Para- / type [p, rounded label with yellow margins] // INDIA / Lyallpur, Punjab / ix.1936 / R. Nath / Ex eggs of *Macropes excavatus* [p] // *Nardo cumaeus*, / Nixon / Paratype 1938 // Pres.by / Imp. Inst. Ent. / B.M.1939-222. [p]’ (Fig. 16).

Current status. Valid as *Baeoneurella cumaea* (Nixon, 1938) (POPOVICI et al. 2018).

Distribution. Oriental, known from the type locality in India and from Pakistan (OHIO STATE UNIVERSITY 2025).

Nardo phaeax Nixon, 1938

(Fig. 17)

Nardo phaeax Nixon, 1938: 283 (original description).

Three paratypes are deposited in the NMPC (type collection):

PARATYPES (3 ♀♀): ‘Para- / type [p, rounded label with yellow margins] // INDIA / Lyallpur, Punjab / ix.1936 / R. Nath / Ex eggs of *Macropes excavatus* [p] // *Nardo phaeax*, / Nixon / Paratype 1938 [p] // Pres.by / Imp. Inst. Ent. / B.M.1939-222. [p]’ (Fig. 17).

Current status. Valid as *Baeoneurella phaeax* (Nixon, 1938) (POPOVICI et al. 2018).

Distribution. Oriental, known from the type locality in India and from Pakistan (OHIO STATE UNIVERSITY 2025).

Oreiscelio turneri Nixon, 1933

(Fig. 18)

Oreiscelio turneri Nixon 1933: 294, text fig. 1 (original description).

Two paratypes are deposited in the NMPC (type collection):

PARATYPES (2 ♀♀): ‘Para- / type [p, rounded label with yellow margins] // Port St. John, / Pondoland. / May. 1924 [p] // S. Africa. / R. E. Turner. / Brit. Mus. / 1924–289. [p] / *Oriscelio / turneri*, Nixon / Paratype 1933. [p]’ (Fig. 18).

Current status. Valid species (NIXON 1933, TALAMAS et al. 2009).

Distribution. Afrotropical, known from Botswana, Kenya, Malawi, Mozambique, Namibia, Somalia, South Africa, Tanzania, Zambia and Zimbabwe (TALAMAS et al. 2009).

Paranteris tenuis Nixon, 1933

(Fig. 19)

Paranteris tenuis Nixon, 1933: 556–559, text fig. 10 (original description).

One paratype is deposited in NMPC (general collection):

PARATYPE (♂): ‘Para- / type [p, round label in yellow frame] // Cape Province: / Somerset East. / November 1930. [p] // S.Africa. / R.E.Turner. / Brit. Mus. / 1930-593. [p] // *Paranteris / tenuis* Nixon. / Paratype 1933. [p]’ (Fig. 19).

Current status. Valid as *Paridris tenuis* (Nixon, 1933) (MASNER 1965).

Distribution. Afrotropical, known from Burkina Faso, Cameroon, Gambia, Ghana, Ivory Coast, Kenya, Nigeria, Sierra Leone, South Africa, Swaziland, Tanzania, Uganda and Zimbabwe (TALAMAS et al. 2013).

Paratrimorus pinguis Nixon, 1933

(Figs 20A–C)

Paratrimorus pinguis Nixon, 1933: 313–314, text fig. 6 (original description).

Psilanteris pinguis (Nixon, 1933), **comb. nov.**

One paratype is deposited in NMPC (general collection):

PARATYPE (♂): ‘Para- / type [p, round label in yellow frame] // Port St. John, / Pondoland. / Dec. 1923. [p] // S. Africa. / R. E. Turner. / Brit. Mus. / 1924–54 [p] // *Paratrimorus / pinguis* Nixon. / Paratype 1933. [p] // *Psilanteris pinguis* (Nixon, 1933) / Talamas E. Det. 2025 [p]’ (Figs 20A–C).

Generic transfer. KOZLOV (1971) treated *Paratrimorus* Kieffer, 1910 as a junior synonym of *Anteris* Förster, 1856, implicitly transferring *Paratrimorus pinguis*. Examination of the paratype in NMPC revealed that this species does not belong in *Anteris* and instead belongs in *Psilanteris* Kieffer, 1916 based on the concept provided in MASNER (1976). Characters that support this transfer are the presence of a skaphion (Fig. 20B), a short metascutellar spine (Fig. 20A),

and the distinctive venation of the forewing, with a very short marginal vein and no postmarginal vein (Fig. 20C). **Current status.** Valid as *Psilanteris pinguis* (Nixon, 1933) comb. nov.

Distribution. Afrotropical, known from South Africa (NIXON 1933).

Procacus dubius Nixon, 1931

(Fig. 21)

Procacus dubius Nixon, 1931: 365–367 (original description).

The paratype is deposited in NMPC (general collection):

PARATYPE (♀): 'Para- / type [p, rounded label in yellow frame] // Aliwal North, / Cape Province. / Dec. 1922. [p] // S. Africa. / R. E. Turner. / Brit. Mus. / 1923–45. [p] // *Procacus / dubius*, Nixon. / Paratype 1931. [p]' (Fig. 21).

Current status. Valid as *Probaryconus dubius* (Nixon, 1931) (MASNER 1965).

Distribution. Afrotropical, known from the type locality in South Africa (NIXON 1931).

Thelepte serapis Nixon, 1931

(Fig. 22)

Thelepte serapis Nixon, 1931: 380–382, fig. 8 (original description).

One paratype is deposited in NMPC (general collection):

PARATYPE (♂): 'Para- / type [p, rounded label in yellow frame] // Port St. John, / Pondoland. / 6-25.Feb.1924. [p] // S. Africa. / R.E.Turner. / Brit. Mus. / 1924–136. [p] // *Thelepte / serapis*, / Nixon. / Paratype 1931. [p]' (Fig. 22).

Current status. Valid as *Leptoteleia serapis* (Nixon, 1931) (MASNER 1976).

Distribution. Afrotropical, known from the type locality in South Africa (NIXON 1931, MASNER 1976).

Vardhana selene Nixon, 1933

(Fig. 23)

Vardhana selene Nixon, 1933: 320, text fig. 7 (original description).

One paratype is deposited in NMPC (type collection):

PARATYPE (♂): 'Para- / type [p, rounded label with yellow margins] // Port St. John, / Pondoland. / 1–17.Mar.1924. [p] // S. Africa. / R.E.Turner. / Brit. Mus. / 1924–177. [p] // *Vardhana / selene* Nixon / Paratype 1933 [p]' (Fig. 23).

Current status. Valid as *Opisthacantha selene* (Nixon, 1933) (MASNER 1976).

Distribution. Afrotropical, known from South Africa (NIXON 1933, VAN NOORT 2025).

Subfamily Teleasinae Ashmead, 1893

Trimorus bohemicus Masner, 1962

(Fig. 24)

Trimorus bohemicus Masner, 1962: 109–111, fig. 3 (original description).

The holotype is deposited in NMPC (general collection):

HOLOTYPE (♀): 'Cis. Les: Kamenné / 19.vii.51. Bčk. [hw, blue ink] // Holotypus [hw, red label] // *Trimorus* ♀ / *bohemicus* Masner [hw] / L. Masner det., 19[p]62[hw]' (Fig. 24).

Current status. Valid species (SZABÓ 1966, KOZLOV 1978).

Distribution. Palearctic, known from Albania, Austria,

Belgium, Bosnia and Herzegovina, Bulgaria, England, France, Greece, Hungary, Moldova, Russia, Sweden and Ukraine (PETROV 2012).

Subfamily Telenominae Thomson, 1861

Asolcus ghorfi Delucchi & Voegelé, 1961

(Fig. 25)

Asolcus ghorfi Delucchi & Voegelé, 1961: 37–39 (original description).

Three paratypes are deposited in NMPC (general collection):

PARATYPES (1 ♂, 2 ♀♀): 'Asolcus / ghorfi n.sp [hw] / V. Delucchi det. [p] // ex *Aelia cognata* / Maroc, Metenés [hw] / V. Deluchi leg. [p] // PARATYPE [p, red label]' (Fig. 25).

Current status. Junior synonym of *Trissolcus scutellaris* (Thomson, 1861) (TALAMAS et al. 2017).

Distribution. Palearctic, known from Austria, Bulgaria, Croatia, France, Georgia, Germany, Greece, Hungary, Iran, Italy, Macedonia, Morocco, Portugal, Russia, South Korea, Spain, Sweden, Syria, Turkey and Turkmenistan (TALAMAS et al. 2017).

Platytenomus danubialis Széleányi, 1939

(Fig. 26)

Platytenomus danubialis Széleányi, 1939: 125–127, fig. 3 (original description).

The allotype is deposited in NMPC (type collection):

ALLOTYPE (♂): 'Bohemia c. / Koda V.53. / Obenberger [hw] // Mus. Nat. Pragae / Inv. [p] 3101 [hw] // AT [hw, red label]' (Fig. 26).

Current status. Valid as *Telenomus danubialis* (Széleányi, 1939). JOHNSON (1992) referenced JOHNSON (1984) for the generic transfer of this species, but we found no mention of *danubialis* in that publication. JOHNSON (1988) treated *Platytenomus* Dodd, 1914 as a junior synonym of *Telenomus* Haliday, 1833 implicitly transferring species from the former to the latter, so making it likely that the erroneous reference is the result of a typographical error.

Distribution. Palearctic, known from the Czech Republic, France, Hungary, Italy, Japan, Russia, Spain, Sweden and United Kingdom (SZÉLEÁNYI 1939, MASNER 1955, FERGUSON 1983, RYU & HIRASHIMA 1989).

Platytenomus hylas Nixon, 1935

(Fig. 27)

Platytenomus hylas Nixon, 1935: 74 (original description).

Five paratypes are deposited in NMPC (general collection):

PARATYPES (5 ♂♂): 'Para- / type [p, rounded label with yellow frame] // Sudan Govt. [p, vertically written] / Shendi / I. W. Cowland / Dec. 1929–Jan.1930 / Bred from eggs of / *Sesamia cretica* [hw] // Ent. Coll. [p] / C8203 [hw] // *Platytenomus / hylas* Nixon / Paratype 1935. [p] // Pres. By / Imp.Inst.Ent. / B.M.1936–224 [p]' (Fig. 27).

Current status. Junior synonym of *Telenomus busseolae* Gahan, 1922 (FERGUSON 1983, JOHNSON 1988).

Distribution. Known from Bangladesh, Cameroon, Egypt, Ghana, Greece, Indonesia, Iran, Iraq, Israel, Kenya, Mauritius, Nigeria, Reunion, Senegal, South Africa, Sudan, Uganda (NIXON 1935, KOZLOV & KONONOVA 1983, POLASZEK & KIMANI 1990, GHAHARI &

BUHL 2015, OHIO STATE UNIVERSITY 2025).

Nirupama auge Nixon, 1935

(Fig. 28)

Nirupama auge Nixon, 1935: 94 (original description).

One paratype is deposited in the NMPC (type collection).

PARATYPE (♀): 'Para- / type [p, rounded label with yellow margins] // Ex. / Hemipteron [p] // GOLD COAST / Afwerase / 31.i.1922 / W.H. Patterson [p] // *Nirupama / auge* Nixon [p] / ♀ [hw] Paratype 1935. [p] // Pres.by / Imp. Inst. Ent. / B.M.1936-224 [p]' (Fig. 27).

Current status. Valid species (NIXON 1935, MASNER 1965, JOHNSON 1992).

Distribution. Afrotropical, known from the type locality in Ghana (NIXON 1935, MASNER 1965).

Acknowledgements

This work was supported by the Ministry of Culture of the Czech Republic IP DKRVO 2024–2028/5.I.b, 00023272. Elijah Talamas was supported by the Florida Department of Agriculture and Consumer Services, Division of Plant Industry. We would like to express our gratitude to Dr. Lubomír Masner (Agriculture and Agri-Food Canada, Canadian National Collection of Insects) for his expert help in the identification and correct taxonomic placement of the type material. Our thanks also go to the Editor-in-Chief, Dr. Petr Kment, for his valuable comments that helped to finalize the manuscript.

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Appendix: Figures 1–28



Fig. 1. Habitus of *Aphanogmus annulicornis* Jansson, 1957, paratype, junior synonym of *Aphanogmus compressus* (Ratzeburg, 1852), with labels.



Fig. 2. Habitus of *Elysocephron hungaricus* Szelenyi, 1936, allotype, with labels.



Fig. 3. Habitus of *Sinaris roubali* Gregor, 1939, holotype, valid as *Sinaris xanthothorax* Szelenyi, 1936, with labels.



Fig. 4. Habitus of *Inostemma staryi* Masner, 1955, holotype, with labels.



Fig. 5. Habitus of *Fidiobia hofferi* Kozlov, 1978, lectotype, with labels.



Fig. 6. Habitus of *Iphitrachelus gracilis* Masner, 1957, holotype, with labels.



Fig. 7. Habitus of *Serphus gravidator antennalis* Tomšik, 1944, holotype, with labels.



Fig. 8. Habitus of *Serphus gravidator niger* Tomšik, 1944, holotype, with labels.



Fig. 9. Habitus of *Breviscelio crenatus* Sundholm, 1970, paratype, valid as *Gryon crenatum* (Sundholm, 1970), with labels.



Fig. 10. Dorsal and lateral habitus of *Dicroscelio leucosius* (Nixon, 1931) comb. nov., paratype, with labels.



Fig. 11. Habitus of *Duta tenuicornis fragilis* Nixon, 1933, paratype, with labels.



Fig. 12. Habitus of *Echthrodesis lamorali* Masner, 1968, paratype, with labels.



Fig. 13. Habitus of *Govinda nona* Nixon, 1933, paratype, valid as *Dicroscelio nonus* (Nixon, 1933), with labels.



Fig. 14. Habitus of *Hadronotus lymantriae* Masner, 1958, holotype, valid as *Gryon lymantriae* (Masner, 1958), with labels.



Fig. 15. Habitus of *Macroteleia unicolor* Nixon, 1931, paratype, valid with the replacement name *Macroteleia nixonii* Masner, 1965, with labels.



Fig. 16. Habitus of *Nardo cumaeus* Nixon, 1938, paratype, valid as *Baeoneurella cumaea* (Nixon, 1938), with labels.



Fig. 17. Habitus of *Nardo phaeax* Nixon, 1938, paratype, valid as *Baeoneurella phaeax* (Nixon, 1938), with labels.



Fig. 18. Habitus of *Oreiscelio turneri* Nixon, 1933, paratype, with labels.



Fig. 19. Habitus of *Paranteris tenuis* Nixon, 1933, paratype, valid as *Paridris tenuis* (Nixon, 1933), with labels.

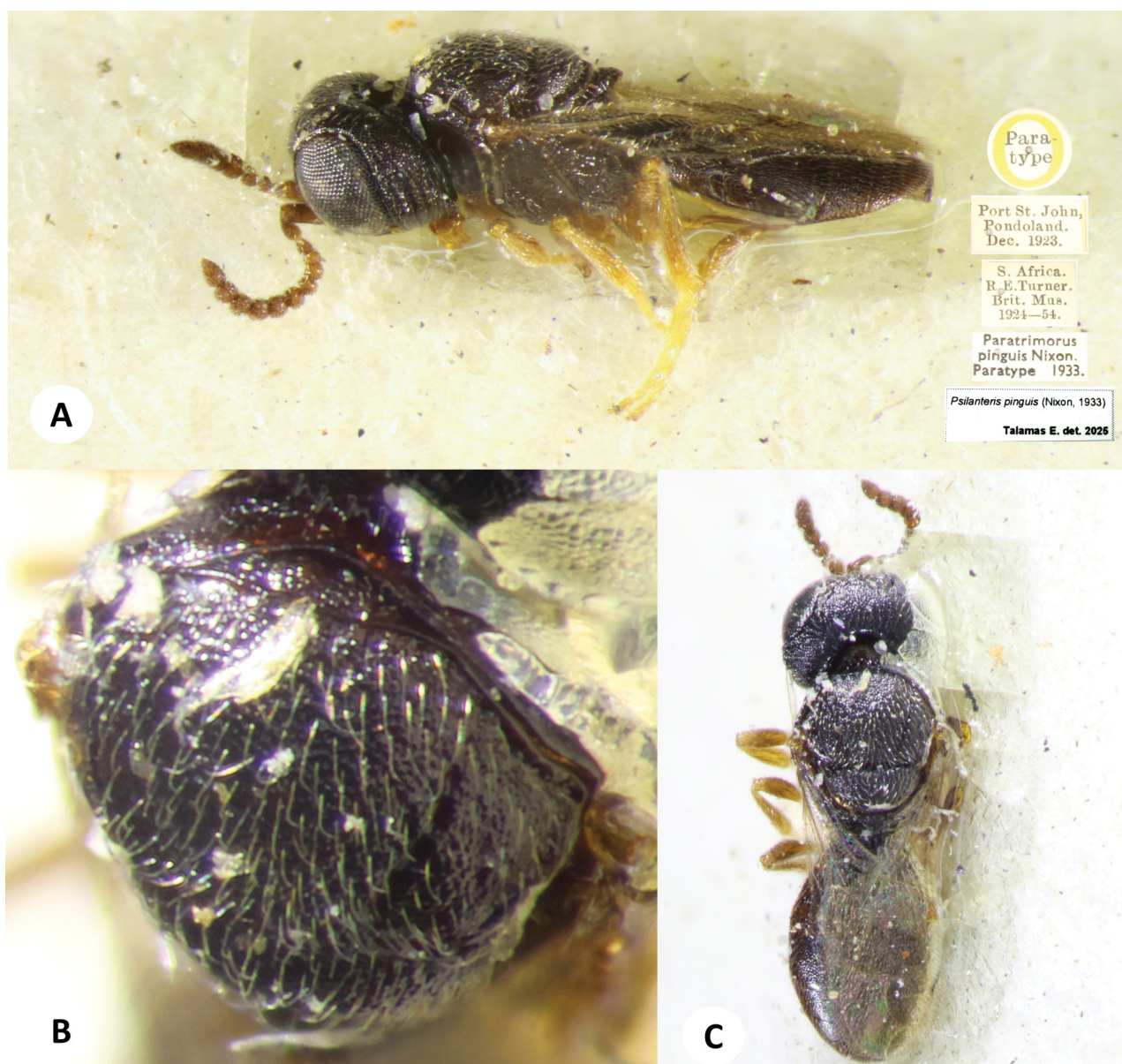


Fig. 20. Paratype of *Psilanteris pinguis* (Nixon, 1933) comb. nov. A – lateral habitus and labels, B – mesonotum anteriorly with skaphion, C – dorsal habitus.



Fig. 21. Habitus of *Procacis dubius* Nixon, 1931, paratype, valid as *Probaryconus dubius* (Nixon, 1931), with labels.



Fig. 22. Habitus of *Thelepte serapis* Nixon, 1931, paratype, valid as *Leptoteleia serapis* (Nixon, 1931), with labels.



Fig. 23. Habitus of *Vardhana selene* Nixon, 1933, paratype, valid as *Opisthacantha selene* (Nixon, 1933), with labels.



Fig. 24. Habitus of *Trimorus bohemicus* Masner, 1962, holotype, with labels.



Fig. 25. Habitus of *Asolcus ghorfii* Delucchi & Voegelé, 1961, paratypes, valid as *Trissolcus scutellaris* (Thomson, 1861), with labels.



Fig. 26. Habitus of *Platytelenomus danubialis* Szelényi, 1939, allotype, valid as *Telenomus danubialis* (Szelényi, 1939), with labels.



Fig. 27. Habitus of *Platytelenomus hylas* Nixon, 1935, paratype, valid as *Telenomus busseolae* Gahan, 1922, with labels.



Fig. 28. Habitus of *Nirupama auge* Nixon, 1935 paratype, with labels.

